

Interactions of polyamines in the measurement of free magnesium concentration by mag-fura-2 and ^{31}P -NMR

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Abstract

Polyamines, particularly spermine, in physiological concentrations interact with mag-fura-2 and the mag-fura-2/ Mg^{2+} complex, resulting in reduced values of free Mg^{2+} concentration. Similarly, polyamines interact with ATP and MgATP . Thus, free Mg^{2+} concentration, as measured by ^{31}P -NMR or mag-fura-2, is underestimated in the presence of polyamines, particularly of spermine.

Key words: Intracellular free magnesium; Magnesium ion, intracellular; Polyamine; Mag-fura-2; NMR, ^{31}P -

1. Introduction

The concentration of intracellular free Mg^{2+} ($[\text{Mg}^{2+}]_i$) has been measured for 25 years with different methods in various cell types. These measurements were done by means of Mg^{2+} -dependent enzymes, Mg^{2+} -sensitive electrodes, null-point method, ^{31}P -NMR, and Mg^{2+} -binding fluorescent dyes [1,2]. The two latter methods are now predominantly being used. In these methods, Mg^{2+} is bound to negatively charged mag-fura-2, changing its fluorescence, or Mg^{2+} is bound to ATP^{4-} , changing its ^{31}P -NMR spectrum. Both methods require specificity of Mg^{2+} binding to mag-fura-2 or ATP^{4-} . However, it is well known that polyamines have similar binding properties to negatively charged ligands as Mg^{2+} [3,4] and may compete with Mg^{2+} in ATP binding.

At pH 7.5 putrescine, spermidine and spermine are bound to ATP^{4-} with apparent binding constants of 290 M^{-1} , 900 M^{-1} and 9500 M^{-1} . For comparison, the MgATP complex binding constant amounted to $17\,200\text{ M}^{-1}$ when measured by the same method [5]. Moreover, a second spermidine and spermine molecule is bound to ATP with an apparent binding constant of

280 M^{-1} and 2400 M^{-1} . With similar affinity as Mg^{2+} , polyamines are also bound to ADP [5], which was also used to measure $[\text{Mg}^{2+}]_i$ by means of ^{31}P -NMR [6].

Since intracellular concentrations of polyamines are in the millimolar range and fluctuate in response to stimuli (for references, see ref. 5), we investigated the interaction of Mg^{2+} and polyamines with mag-fura-2 and the interaction of polyamines with ATP and MgATP in the estimation of free Mg^{2+} by ^{31}P -NMR and mag-fura-2.

2. Materials and methods

2.1. Fluorescence spectra

Mag-fura-2 ($0.2\text{ }\mu\text{M}$) was dissolved in K^+ medium containing: 140 mM KCl , 10 mM NaCl , 30 mM Hepes-Tris (pH 7.4). MgCl_2 and polyamines were added as indicated.

Fluorescence spectra were recorded with a Perkin Elmer LS 50 Luminescence Spectrometer. Excitation: $300\text{--}400\text{ nm}$, Emission: 505 nm .

2.2. Measurement of free Mg^{2+} concentration

Mag-fura-2 ($0.2\text{ }\mu\text{M}$) was dissolved in K^+ medium (see above). Every 50 s , 0.2 mM MgCl_2 was added in

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the presence of various concentrations of spermine as indicated.

Fluorescence ratios of the mag-fura-2/ Mg^{2+} complex at excitation 335 and 378 nm and emission 505 nm were measured at 37°C with a Perkin Elmer LS 50 Luminescence Spectrometer using the 'Intracellular Biochemistry' software of the producer. For the mag-fura-2/ Mg^{2+} complex a K_d of 1.5 mM [7] was used.

Maximal fluorescences were measured after addition of 1.2 mM CaCl_2 and minimal fluorescences after addition of 5 mM EDTA.

2.3. ^{31}P -NMR spectra

The solutions contained (in mM): 5 Na_2ATP , or 5 Na_2ATP plus 5 MgCl_2 or 12.5 MgCl_2 without

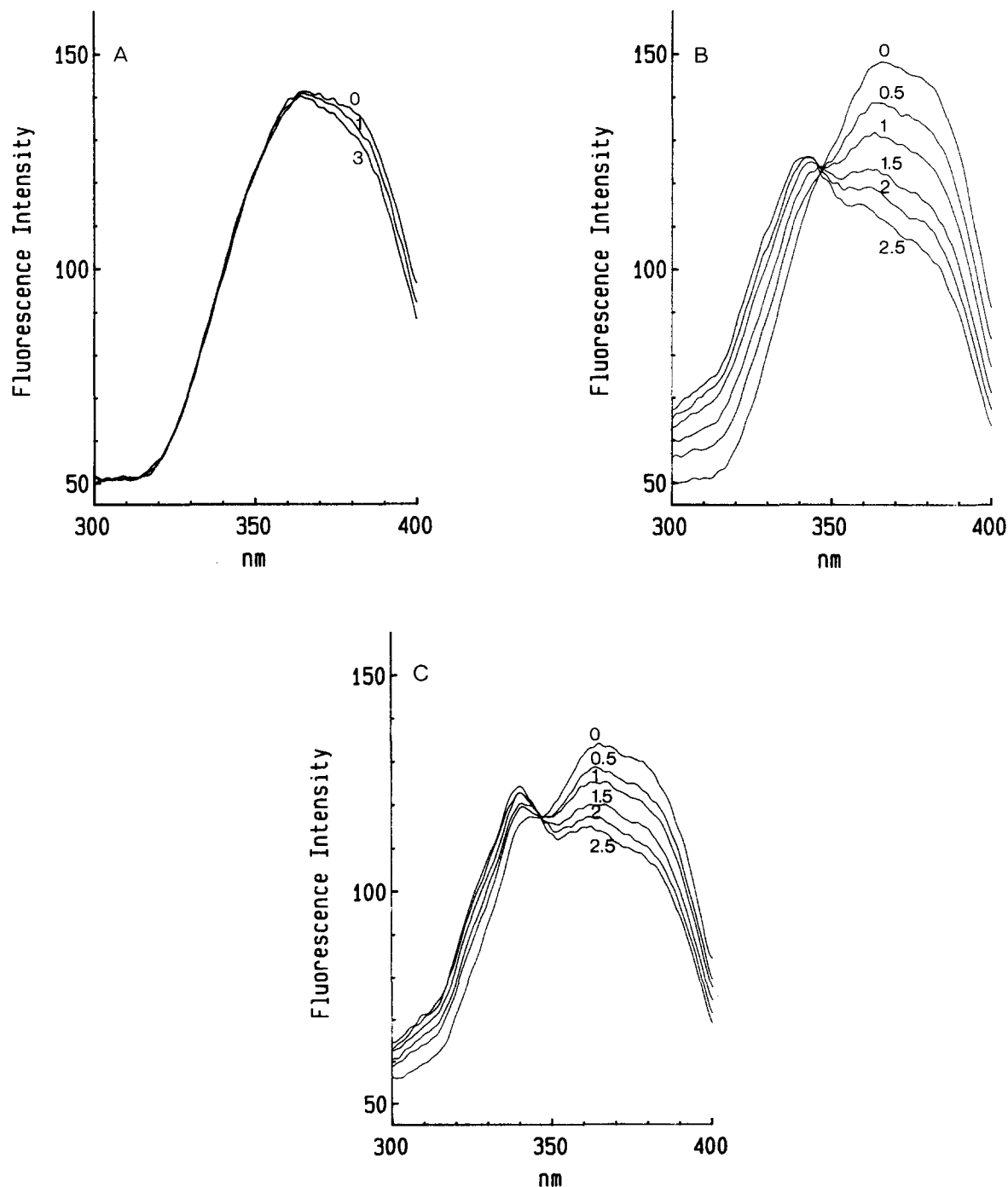


Fig. 1. Fluorescence spectra of mag-fura-2. Mag-fura-2 (0.2 μM) was dissolved in dimethyl sulfoxide and diluted in K^+ medium. (A) Mag-fura-2 plus various spermine concentrations (mM) as indicated; (B) mag-fura-2 plus various MgCl_2 concentrations (mM) as indicated; (C) mag-fura-2 with 3 mM spermine plus various MgCl_2 concentrations (mM) as indicated.

polyamines or with 5 putrescine · 2 HCl, 5 spermidine · 3 HCl or 5 spermine · 4 HCl as indicated, dissolved in 140 KCl, 20 TrisCl (pH 7.2). Na₂ATP was neutralized with Tris. To 2.0 ml of the solutions 0.7 ml D₂O was added to permit heteronuclear field frequency locking on deuterium.

³¹P-NMR measurements were done on a Bruker AM500 spectrometer with a ³¹P-resonance of 202 MHz. All measurements used a one-pulse sequence (90° pulse) without proton decoupling. The probe head temperature was held constant at 310 K and 128 acquisitions were recorded with a pulse repetition delay of 1 s and a sweep width of 20 kHz. Prior to the fast Fourier transformation an exponential multiplication (line broadening factor = 1.2 Hz) of the 64 Kbyte time domain data points was carried out and yielded a digital resolution of 0.31 Hz/point. The resonances were referenced to external H₃PO₄.

Free Mg²⁺ concentration ([Mg²⁺]_{free}) in the solutions was calculated according to Eq. (1) [6,8]:

$$[\text{Mg}^{2+}]_{\text{free}} = K_d^{\text{MgATP}} (\Phi^{-1} - 1) \quad (1)$$

For K_d^{MgATP} (dissociation constant of MgATP) 50 μM was taken [8].

Φ was calculated from the chemical shift differences between the α- and β-phosphoryl group resonances of ATP ($\delta_{\alpha\beta}$), MgATP ($\delta_{\alpha\beta}^{\text{MgATP}}$, Mg²⁺ excess) and shift difference in the presence and absence of polyamines (($\delta_{\alpha\beta}^x$) according to Eq. (2):

$$\Phi = (\delta_{\alpha\beta}^x - \delta_{\alpha\beta}^{\text{MgATP}}) / (\delta_{\alpha\beta}^{\text{ATP}} - \delta_{\alpha\beta}^{\text{MgATP}}) \quad (2)$$

Additionally, [Mg²⁺]_{free} was calculated from the αβ and βγ shift difference according to Eq. (3) [9]:

$$\Phi = \frac{1}{2} \left\{ \frac{(\delta_{\alpha\beta}^x - \delta_{\alpha\beta}^{\text{MgATP}})}{(\delta_{\alpha\beta}^{\text{ATP}} - \delta_{\alpha\beta}^{\text{MgATP}})} + \frac{(\alpha_{\beta\gamma}^x - \delta_{\beta\gamma}^{\text{MgATP}})}{(\delta_{\beta\gamma}^{\text{ATP}} - \delta_{\beta\gamma}^{\text{MgATP}})} \right\} \quad (3)$$

For the calculation of the shift differences, the arithmetic means of the α (α₁, α₂) and γ (γ₁, γ₂) peaks and the β₂ and β peak respectively (in ppm) were used.

Furthermore, [Mg²⁺]_{free} of the equimolar MgATP solution was calculated according to Eq. (4):

$$[\text{Mg}^{2+}]_{\text{free}} = -\frac{1}{2}([\text{ATP}]_{\text{tot}} - [\text{Mg}^{2+}]_{\text{tot}} + K_D^{\text{MgATP}}) + \left(\frac{1}{4}([\text{ATP}]_{\text{tot}} - [\text{Mg}^{2+}]_{\text{tot}} + K_D^{\text{MgATP}})^2 + [\text{Mg}^{2+}]_{\text{tot}} \cdot K_D^{\text{MgATP}} \right)^{1/2} \quad (4)$$

([ATP]_{tot} = total ATP concentration, [Mg²⁺]_{tot} = total Mg²⁺ concentration).

3. Results

3.1. Interaction of polyamines with mag-fura-2 and Mg²⁺/mag-fura-2

As can be seen from Fig. 1A, spermine alone had only a small effect on the fluorescence of mag-fura-2 in the range of 378 nm and 335 nm, which are used to determine [Mg²⁺]_{free} [6,7]. Putrescine and spermidine did not have any significant effect on mag-fura-2 fluorescence either alone or in the presence of Mg²⁺ (not shown). However, the fluorescence of the Mg²⁺/mag-fura-2 complex (Fig. 1B) was considerably changed in the presence of spermine (Fig. 1C). The interaction of spermine in the fluorescence of mag-fura-2 and Mg²⁺/mag-fura-2 was caused by reaction of spermine with mag-fura-2, since spermine itself did not fluoresce or absorb at the wavelengths where measurements were done (data not shown).

To ascertain the interaction of spermine with the Mg²⁺/mag-fura-2 complex quantitatively, every 50 s 0.2 mM MgCl₂ was added to a solution of mag-fura-2 in K⁺ medium (pH 7.4) and the concentration of free Mg²⁺ was measured. As shown in Fig. 2 (curve A), in the absence of spermine correct values of [Mg²⁺]_{free} were obtained as expected. However, in the presence of 0.5 mM spermine (curve B), 30% and in the presence of 3 mM spermine (curve C) 50% lower values of [Mg²⁺]_{free} were determined.

3.2. Interaction of polyamines with ATP and MgATP in ³¹P-NMR

As can be seen from Table 1A, there was only a slight interaction of the polyamines with ATP, according to the series putrescine < spermidine < spermine.

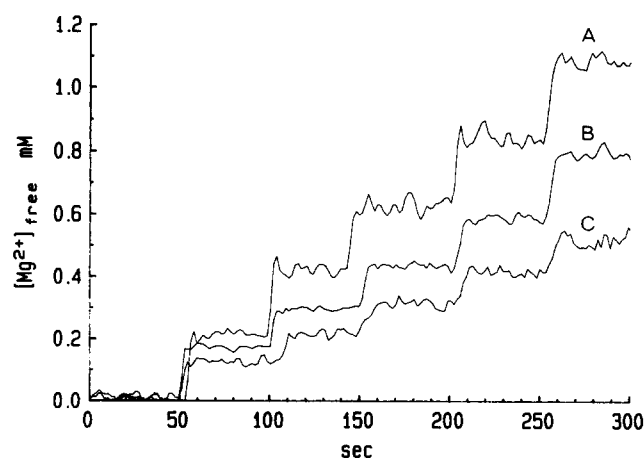


Fig. 2. Effect of spermine on [Mg²⁺]_{free}. Mag-fura-2 (0.2 μM) was dissolved in dimethyl sulfoxide and diluted in K⁺ medium. Every 50 s 0.2 mM MgCl₂ were added. (A) In the absence of spermine; (B) in the presence of 0.5 mM spermine; (C) in the presence of 3 mM spermine.

Table 1A

Effect of polyamines (3.7 mM) on ^{31}P -resonances of ATP (3.7 mM)

| ^{31}P -ATP resonance | ATP | MgATP * | ATP + putrescine | ATP + spermidine | ATP + spermine |
|--------------------------------|--------|---------|------------------|------------------|----------------|
| γ_1^{**} | -6.59 | -5.25 | -6.59 | -6.58 | -6.52 |
| γ_2 | -6.68 | -5.33 | -6.68 | -6.68 | -6.62 |
| α_1 | -10.80 | -10.40 | -10.81 | -10.82 | -10.84 |
| α_2 | -10.90 | -10.48 | -10.91 | -10.92 | -10.94 |
| β_1 | -21.63 | -18.67 | -21.64 | -21.67 | -21.66 |
| β_2 | -21.73 | -18.75 | -21.73 | -21.75 | -21.75 |
| β_3 | -21.83 | -18.83 | -21.83 | -21.84 | -21.85 |

Values in ppm. Mean of two experiments.

* MgATP, 9.25 mM Mg^{2+} plus 3.7 mM ATP. ** γ_1 , γ_2 , α_1 , α_2 , β_1 , β_2 , β_3 mean α and γ doublets and β triplets of ^{31}P -ATP resonances.

Table 1B

Effect of polyamines (3.7 mM) on ^{31}P -resonances of MgATP ($[\text{Mg}^{2+}]_{\text{tot}} = 3.7 \text{ mM}$; $[\text{ATP}]_{\text{tot}} = 3.7 \text{ mM}$)

| ^{31}P -ATP resonance | MgATP | MgATP + putrescine | MgATP + spermidine | MgATP + spermine |
|--------------------------------|--------|--------------------|--------------------|------------------|
| γ_1 | -5.24 | -5.26 | -5.34 | -5.44 |
| γ_2 | -5.32 | -5.34 | -5.42 | -5.52 |
| α_1 | -10.34 | -10.35 | -10.40 | -10.46 |
| α_2 | -10.42 | -10.43 | -10.47 | -10.54 |
| β | -18.92 | -18.96 | -19.07 | -19.33 |

Values in ppm. Mean of two experiments

The interaction of Mg^{2+} with ATP was particularly seen from the shift of the β peak (Table 1B). In the presence of equimolar Mg^{2+} and ATP concentrations the β peak is broadened which is due to Mg^{2+} exchange between MgATP and ATP [10]. Hence β_1 , β_2 and β_3 could not be differentiated. When Mg^{2+} was in excess to ATP, again 3 β peaks occurred (Table 1A).

The polyamines interacted with MgATP. The interaction between polyamines and MgATP again corresponded to the series putrescine < spermidine < spermine (Table 1B) which is in agreement with their ATP complex binding constants [5].

For a quantitative analysis of the interaction with MgATP in ^{31}P -NMR, the concentrations of free Mg^{2+} were calculated according to the above given Eq. (1).

Table 2

Concentration of free Mg^{2+} ($[\text{Mg}^{2+}]_{\text{free}}$) in solutions with 3.7 mM MgCl_2 and 3.7 mM ATP in the absence and presence of 3.7 mM polyamines measured by ^{31}P -NMR

| Polyamine | $[\text{Mg}^{2+}]_{\text{free}}$ (mM) (Eqs. (1), (2)) | $[\text{Mg}^{2+}]_{\text{free}}$ (mM) (Eqs. (1), (3)) |
|------------|--|--|
| – | 0.509 | 0.455 |
| Putrescine | 0.444 | 0.395 |
| Spermidine | 0.345 | 0.325 |
| Spermine | 0.197 | 0.177 |

$[\text{Mg}^{2+}]_{\text{f}}$ was calculated according to Eqs. (1), (2) and (3) (see Section 2).

Table 3

Concentration of free Mg^{2+} ($[\text{Mg}^{2+}]_{\text{free}}$) in solutions with 3.7 mM MgCl_2 and 3.7 mM ATP in the absence and presence of 3.7 mM polyamines measured at 37°C by mag-fura-2 (see Section 2)

| Polyamine | $[\text{Mg}^{2+}]_{\text{free}}$ (mM) |
|------------|---------------------------------------|
| – | 0.51 ± 0.01 |
| Putrescine | 0.44 ± 0.01 |
| Spermidine | 0.35 ± 0.02 |
| Spermine | 0.24 ± 0.01 |

Mean \pm S.E.M. of three experiments.

$[\text{Mg}^{2+}]_{\text{free}}$ calculated according to Eq. (4) amounted to 0.41 mM. Table 2 shows that the values of $[\text{Mg}^{2+}]_{\text{free}}$ determined by ^{31}P -NMR are in agreement with the value of $[\text{Mg}^{2+}]_{\text{free}}$ calculated according to Eq. (4). The values of $[\text{Mg}^{2+}]_{\text{free}}$ were reduced by the polyamines according to the series putrescine < spermidine < spermine.

3.3. Effect of polyamines on $[\text{Mg}^{2+}]_{\text{free}}$ of MgATP solutions measured by mag-fura-2

In order to compare the effects of polyamines on the measurement of $[\text{Mg}^{2+}]_{\text{free}}$ by ^{31}P -NMR, in analogous experiments $[\text{Mg}^{2+}]_{\text{free}}$ of MgATP solutions was measured by mag-fura-2. As shown in Table 3, measurement of $[\text{Mg}^{2+}]_{\text{free}}$ by mag-fura-2 yielded the same values of $[\text{Mg}^{2+}]_{\text{free}}$ as measured by ^{31}P -NMR. $[\text{Mg}^{2+}]_{\text{free}}$ measured by mag-fura-2 was similarly reduced as when measured by ^{31}P -NMR.

4. Discussion

During the last few years mag-fura-2 and ^{31}P -NMR were the most frequently used methods to measure $[\text{Mg}^{2+}]_{\text{f}}$. The interaction of Ca^{2+} and H^{+} were considered in both methods [6,8]. The interaction of polyamines, particularly of spermine, has not yet been taken into account in these measurements.

The intracellular content of polyamines amounts to 1–5 mmol spermine/kg wet weight for liver, pancreas and prostate and to 8–9 mmol spermidine/kg wet weight for prostate and pancreas [3,4,11,12] and correspond to total intracellular Mg^{2+} content. Cellular polyamines are compartmentalized, and a part of the polyamines within the various compartments is bound to negatively charged ligands [3,4,13,14]. As yet there is no method available to determine the concentration of free polyamines besides bound polyamines. The complex binding constants of spermine are about half of the Mg^{2+} complex binding constants [5]. Therefore, in analogy to intracellular Mg^{2+} , it may be supposed that approximately 20% of total spermine is free. For putrescine and spermidine the rate of free polyamine may be higher than for spermine because of their lower

complex binding constants [5]. In Figs. 1 and 2 the used spermine concentrations varied between 0.5 and 3 mM and in Tables 2 and 3 the used polyamine concentrations amounted to 3.7 mM and may be within or near the physiological range in some cell types. Thus, $[Mg^{2+}]_i$ measured by these two methods is underestimated in the presence of polyamines, particularly in the presence of spermine. This effect must be considered when values for $[Mg^{2+}]_i$ measured by mag-fura-2 and ^{31}P -NMR are compared with those measured by the null-point method or Mg^{2+} -sensitive electrodes.

The mechanism by which polyamines reduce the measured values of $[Mg^{2+}]_{free}$ can be attributed to competition of polyamines with Mg^{2+} for common binding sites of mag-fura-2 and ATP^{4-} , and to lower responses in fluorescence or ^{31}P -NMR of the polyamine-mag-fura-2 and polyamine-ATP complex than the corresponding Mg^{2+} complexes. The alternative mechanism may be binding of Mg^{2+} to polyamines and thus reducing $[Mg^{2+}]_{free}$. When Mg^{2+} binding to spermine was investigated by proton magnetic resonance, it was found that most of the spermine existed in a free form [15]. This result implies that there is no significant binding of Mg^{2+} to spermine. Hence, reduction of measured $[Mg^{2+}]_{free}$ by binding of Mg^{2+} to polyamines can be neglected. Also, an association constant for Mg^{2+} binding to spermine is not available.

Using mag-fura-2, it was found that some cell types showed an increase in $[Mg^{2+}]_i$ after stimulation with various effectors, such as epidermal growth factor [16,17], insulin [16], vasopressin [18], endothelin [18], carbachol [19], cGMP [20] and atrial natriuretic peptide [20]. An increase in $[Mg^{2+}]_i$ was also found by isoproterenol in heart muscle cells as measured by ^{31}P -NMR [21]. A decrease of $[Mg^{2+}]_i$ was induced by cAMP, PTH and calcitonin [20]. From these results it was concluded that $[Mg^{2+}]_i$ may play a role in the intracellular mechanisms of these effectors. In these experiments the alterations of $[Mg^{2+}]_i$ were very small and amounted to 0.1 to 0.2 mM. However, the concentration of polyamines (besides Ca^{2+} or H^+) is also changed by some of these effectors [3,13,14,22].

Hence, the interaction of polyamines in the measurement of $[Mg^{2+}]_i$ cannot be defined quantitatively

and the real alterations in $[Mg^{2+}]_i$ by effectors remain uncertain.

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